

Supplementary Information

Multiple scales of diversification within natural populations of archaea in hydrothermal chimney biofilms

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Sample descriptions

Carbonate chimney samples LC1408 (full sample name 3881-1408), LC1404 (3869-1404), and LC1443 (3869-1443) were collected from the Lost City Hydrothermal Field (LCHF, depth, ~735 m; latitude, 30.12; longitude, -42.12) with DSV Alvin during cruise AT07-34 aboard the R/V Atlantis in April/May 2003 (<http://www.lostcity.washington.edu>). Sample LC0424 (H03_072705_R0424) was collected by DSV *Hercules* during the 2005 Lost City Expedition aboard the R/V *Ronald H. Brown*. LC0424 and LC1408 were collected from a site known as Marker 3 or 'Poseidon,' a 60 m tall edifice emitting fluids at temperatures ranging from 55-88°C (Kelley *et al.* 2005). LC1408 minerals appeared bright white in color, very friable, and not lithified. Samples LC1404 and LC1443 are from a structure named Marker C, a ~50 cm wide flange structure with several small (centimeters tall) chimneys growing on the top of the flange. LC1404 was collected from the front of the flange, and LC1443 was a small spire collected from the top. Both samples were cream white with a reddish discoloration that remains unexplained (Ludwig *et al.*, 2006). Additional published characteristics of the samples are summarized in Table S1.

Shipboard, subsamples of chimney material were frozen immediately at -80°C and remained frozen until onshore analysis. DNA was extracted from carbonate chimney samples according to a protocol

modified from previous reports (Brazelton *et al.*, 2006; Barton *et al.* 2006) and summarized here. After crushing a frozen carbonate sample with a sterile mortar and pestle, approximately 0.25 – 0.5 g of chimney material were placed in a 2 mL microcentrifuge tube containing 250 μ L of 2x buffer AE (200 mM Tris, 50 mM EDTA, 300 mM EGTA, 200 mM NaCl, pH 8) and 2 μ g of poly-dIdC (Sigma-Aldrich) and incubated at 4°C overnight to allow chelation of salts and binding of DNA to poly-dIdC. Between 36-72 replicate tubes were processed in parallel, and approximately 15 g of carbonate minerals were processed for each sample. Proteinase K (final concentration 1.2 mg/mL) and 10 μ L of 20% SDS were added to each tube before incubation at 37°C for at most 30 min. A further 150 μ L of 20% SDS and 500 μ L of phenol:chloroform:isoamyl alcohol (25:24:1 ratio by volume) were added to each tube before centrifugation at 12,000 g for 10 min. Supernatants were transferred to clean tubes for a second phenol:chloroform:isoamyl alcohol extraction. After centrifugation, supernatants were pooled into SnakeSkin dialysis tubing (Pierce) and dialyzed against 20 mM EGTA overnight at 4°C. This large scale dialysis step proved to be very efficient in removing inorganic minerals and organic inhibitors. After dialysis, DNA was precipitated by adding 0.1 vol 3M sodium acetate and 1 vol isopropanol and stored at -20°C for 2-4 hours. Pellets were collected by centrifugation at 16,000g for 20 min at 8°C, washed once in 70% ethanol, dried in a vacuum centrifuge, and resuspended in TE (10 mM Tris, 1mM EDTA, pH 8). Typical yield was ~35 mg of DNA per g of carbonate chimney material.

Construction and sequencing of clone libraries

Two 16S rRNA clone libraries including a total of 486 clones (GenBank accession numbers FJ791302-FJ791787) from sample LC0424 were constructed by the DOE Joint Genome Institute according to the standard protocol published on their website: <http://my.jgi.doe.gov/general/index.html>. The V6-ITS clone libraries including a total of 516 clones from three samples (accession numbers GQ272945-GQ273460) were constructed from amplicons covering the 16S rRNA V6 region downstream through the intergenic transcribed spacer (ITS) region to the 23S rRNA. PCR amplification was conducted

according to the protocol of Huber *et al.* (2006). The forward primer (886F-LCMS: GAAGTACGGCCGCAAGGC) targets a region just upstream of the Lost City Methanosarcinales V6 region, and the reverse primer (58Ra: GCTTATCGCAGCTTGSCACG) targets the 5' end of the archaeal 23S rRNA gene (Huber *et al.* 2006). V6-ITS amplicons were reconditioned using the protocol of Thompson *et al.* (2002) and cloned using the TOPO-TA cloning kit (Invitrogen) according to the manufacturer's instructions. Cloned inserts were sequenced at the University of Washington High-Throughput Genomics Unit (www.htseq.org) with sequencing primers described by Huber *et al.* (2006). Because of inhibitors that could not be removed from the DNA preparations, PCR amplification of V6-ITS clones required 34-38 cycles of PCR amplification. It is possible that the higher evenness in LC1408 (Table 1) resulted from the higher number of cycles (38) used during PCR amplification of this sample compared to other two samples, which required only 34 cycles. The higher diversity in LC1408 and LC1443 compared to LC1404, however, is unlikely to be affected by cycle number or polymerase error, because only 34 cycles were used for both LC1443 and LC1404 and because of the high mutation rates in these libraries compared to that expected from polymerase and sequencing error, as described in the main text. More amplification cycles may have been required for sample LC1408 because it contained 100x lower archaeal density than the other two samples (Table X?) even though efforts were made to equalize DNA template concentrations. All alignments were calculated with MUSCLE (Edgar *et al.*, 2004).

Analysis of tag pyrosequences

Protocols for construction and sequencing of V6 amplicon libraries have been described previously (Sogin *et al.*, 2006; Huber *et al.*, 2007). Tag sequences were screened for quality as recommend by Huse *et al.* (2007). Sequences assigned to the family *Methanosarcinaceae* by GAST (Huse *et al.*, 2008) were aligned with MUSCLE (Edgar *et al.*, 2004). Distance matrices were calculated with quickdist as described by Sogin *et al.* (2006) except that terminal gaps were penalized in our study because we

inspected the 3' ends to confirm that primers were accurately trimmed and that the most common 3' deletions were not the result of incomplete sequences. Evenness values were derived from the Shannon-Weaver index as calculated by DOTUR (Schloss *et al.*, 2005), and 97% sequence similarity OTUs were calculated with DOTUR. To normalize relative abundances of each sequence among samples, tags were randomly resampled down to the sample with the fewest tags (LC1408: 14,869 tags) using Daisy-Chopper (available at <http://www.genomics.ceh.ac.uk/GeneSwytch/Tools.html>).

Community similarities among samples

The abundance distributions of tag sequences in the three samples were highly similar, though sample LC1404 is more similar to LC1443 (94% Bray-Curtis similarity), which was sampled ~20 cm away on the same chimney, than to LC1408 (90% Bray-Curtis similarity), which was collected from a different chimney. After removing the one dominant sequence (because the Bray-Curtis index is weighted toward dominant members) and sequences occurring only once in one sample (to decrease the number of heavily undersampled sequences), the abundance distributions of the 483 remaining sequences (Fig. 2a) yielded a greater Bray-Curtis similarity between samples from the same chimney (LC1404 and LC1443, 79%) than between samples from different chimneys, (70-71%). If only very rare sequences (represented by fewer than 10 tags in each sample after normalization) were considered in the similarity calculation, the same trend was observed: LC1404 and LC1443 were 46% similar but only 35-38% similar to LC1408 according to the Bray-Curtis index. Therefore, the abundances of dominant as well as rare sequences are more similar in samples from the same chimney than in samples from different chimneys.

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Chimney Sample	Chimney Location	Max fluid temp (°C)	Max fluid H ₂ (mmol kg ⁻¹)	Max fluid CH ₄ (mmol kg ⁻¹)	Cells g ⁻¹ dry weight ^a	Archaea ^b	Bacteria ^b	LCMS ^b	Total organic carbon (%)	^{TM13} C _{toc} (‰ vs. VPDB)
LC1408	Marker 3	88	13.26	1.55	2.0 x 10 ⁻⁸	25%	14%	18%	n.d.	n.d.
LC1404	Marker C	70	14.38	1.98	1200 x 10 ⁻⁸	41%	8%	32%	0.20	-7.8
LC1443	Marker C	70	14.38	1.98	1600 x 10 ⁻⁸	38%	10%	21%	n.d.	n.d.

^a Determined by DAPI-staining

^b Percentage of DAPI-stained cells detected by FISH probe specific to each group

Table S1. Previously published characteristics of the three carbonate chimney samples from which V6 tags and V6-ITS clone libraries were sequenced. Fluid temperatures and concentrations of H₂ and CH₄ are maximum values reported by Proskurowski *et al.* (2006 & 2008). Cell densities and proportions of phylogenetic groups are from Schrenk *et al.* (2004) and M. Schrenk (doctoral dissertation, 2005). Organic carbon concentrations and isotopic measurements are from Bradley *et al.* (2009). Fluid temperature and chemistry are identical for samples LC1404 and LC1443 because these carbonate samples were collected from the same chimney.

Figure S1 with caption below:

1

thermoauto_A TACACAAA-- ----AAGA ATAAAG---- ----AGATGTG TGCTTTTCG- ---GGGATTA CTC---CTCC CACTG--TGA TGGGGC----

thermoauto_B TACACAAA-- ----AAGA ATAAAG---- ----AGATGTG TGCTTTTCG- ---GGGATTA CTC---CTCC CACTG--TGA TGGGGC----

thermophila_A TACACAAA-- ----AAGA ATAAAG---- ----AGTGTG CATAA---- ----A TCG---GCCG GAAGC--TGA TAGG-----

thermophila_B TACACAAA-- ----AAGA ATAAAG---- ----AGTGTG CATAA---- ----A TCG---GCCG GAAGC--TGA TAGG-----

LCMS TACACAAA-- ----AAGA ATAAAG---- ----CGATCTG TATAATACAG ATCAACACTA CTA---ATTA GATGT--CGA AAAACTA----

stadtmanae_A AATATAC--- ----AATT AAAGA---- ----TATATTG TTACATTAG TTTAACAGTA TTATTCATTA TTTTAAATAA TAAATTA----

stadtmanae_B AACATATAAA ATTTATAAGT AAAGGATAAT AATTTTCATAT CCAAAAATTTG TGTACA---- --TACACTA TTA---TATG AATTT--TAA TAAGTTATTT

stadtmanae_C AACATATAAA TTTA--TAAT AAAGGATAAT AATTTTCATAT CCAAAAATTTG TGTACA---- --TACACTA TTA---TATG AATTT--TAA TAAGTTATTT

stadtmanae_D AACATATAAA TTTA--TAAT AAAGGATAAT AATTTTATAT CC--AAATTTG TGTACA---- --TACACTA TTA---TATG AATTT--TAA TAAGTTATTT

burtonii_A -----AAGC A----- --AGATCCG CACAAAGCGG ATCACCCTA TCA---GTCA GAAAT--CGA TAAACTG----

burtonii_B -----AAGC A----- --AGATCCG CACAAAGCGG ATCACCCTA TCA---GTCA GAAAT--CGA TAAACTG----

burtonii_C -----AAGC A----- --AGATCCG CACAAAGCGG ATCACCCTA TCA---GTCA GAAAT--CGA TAAACTG----

barkeriA -----AAGC AAAA----- --AAACTCA CCA---CCCA GATGC--CGA TAAACCG----

barkeriB -----AAGC AAAA----- --AAACTCA CCA---CCCA GATGC--CGA TAAACCG----

barkeriC -----AAGC AAAA----- --AAACTCA CCA---CCCA GATGC--CGA TAAACCG----

mazeiA -----AAGC ATAA----- --AACAAATA TCA---CCCA GATGC--CGA TAAACCG----

mazeiB -----AAGC ATAA----- --AACAAATA TCA---CCCA GATGC--CGA TAAACCG----

mazeiC -----AAGC ATAA----- --AACAAATA TCA---CCCA GATGC--CGA TAAACCG----

acetivoransB -----AAGC CGAAAA----- --AACACTA CCA---CCCA GATGC--CGA TAAACCG----

acetivoransA -----AAGC CGAAAA----- --AACACTA TCA---CCCA GATGC--CGA TAAACCG----

acetivoransC -----AAGC CGAAAA----- --AACACTA TCA---CCCA GATGC--CGA TAAACCG----

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thermoauto_A -----ACCTTAACT GT----- TCTGGTTCTA

thermoauto_B -----ACCTTAACT GT----- TCTGGTTCTA

thermophila_A -----TTCGTCCT TGACCTGTTG CTGGGATCTA

thermophila_B -----TTCGTCCT TGACCTGTTG CTGGGATCTA

LCMS -----CCAAG TTCAAGAA-- ---CCATAAGT AAG----- TTGTGATGAA

stadtmanae_A -----GAAATAA TTTTATAGT CAGCTTACAT TTTAACAG-- --ATGATAAGT GAA----- --GAGATGAA

stadtmanae_B TTTCATATTC ATCATATTTT ACAAAACATGT AATTTGTTGT CTTATAGGTG CAGCTTACAT TTTAACAG-- --ATGATAAGT GAA----- --GAGATGAA

stadtmanae_C TTTCATATTC ATCATATTTT ACAAAACATGT AATTTGTTGT CTTATAGGTG CAGCTTACAT TTTAACAG-- --ATGATAAGT GAA----- --GAGATGAA

stadtmanae_D TTTCATATTC ATCATATTTT ACAAAACATGT AATTTGTTGT CTTATAGGTG CAGCTTACAT TTTAACAG-- --ATGATAAGT GAA----- --GAGATGAA

burtonii_A -----CCAAA TTCAACAAC CACTTTCAT GAA----- CTGAGATTGC

burtonii_B -----CCAAA TTCAACAAC CACTTTCAT GAA----- CTGAGATTGC

burtonii_C -----CCAAA TTCAACAAC CACTTTCAT GAA----- CTGAGATTGC

barkeriA -----AACAAA --ATCCTCACC -----A

barkeriB -----AACAAA --ATCCTCACC -----A

barkeriC -----AACAAA --ATCCTCACC C----- CCGAGATCCG

mazeiA -----AACAAA --TCCTCAAAC -----CTGAGATCCA

mazeiB -----AACAA-- --ATCCTCAAA CCAGAAATCG ATAAGATTTT

mazeiC -----AACAA-- --ATCCTCAAA CCAGAAATCG ATAAGATTTT

acetivoransB -----ATCAAAA --TCCTCAAAC -----CTGAGATCCT

acetivoransA -----ATCAA-- --ATCCTCACC -----A

acetivoransC -----ATCAA-- --ATCCTCACC -----A

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thermoauto_A TCTGTATCCT -----

thermoauto_B TCTGTATCCT -----

thermophila_A TTTGG-----

thermophila_B TTTGG-----

LCMS TTTGGGG-----C TTGTAGATCA GTTGGAGAT CGCT-----

stadtmanae_A ATTAATGAT ---ATTCCAT TTATGGAGTA --TTAGAAGAA TAATCATCGA TCATATGTA- TATCCAGTAT AAGCATTAAA ACTAATAGTA TGTTTATTAC

stadtmanae_B ATTAATGAT ---ATTCCAC TTGTGGAGTA TTTAGAAGAA TAATCATCGA TCATATGTAC TATCCAGTAT AAGCATTAAA ACTAATAGTA TGTTTATTAC

stadtmanae_C ATTAATGAT ---ATTCCAT TTATGGAGTA TTTAGAAGAA TAATCATCGA TCATATGTAC TATCCAGTAT AAGCATTAAA ACTAATAGTA TGTTTATTAC

stadtmanae_D ATTAATGAT ---ATTCCAT TTATGGAGTA TTTAGAAGAA TAATCATCGA TCATATGTAC TATCCAGTAT AAGCATTAAA ACTAATAGTA TGTTTATTAC

burtonii_A CTGGAAGTT -----CTCA GTTGGATCAA T-----

burtonii_B CTGGAAGTT -----CTCA GTTGGATCAA T-----

burtonii_C CTGGAAGTT -----CTCA GTTGGATCAA T-----

barkeriA TTTAAATCAT CGATCATAAT CTAATGATCA ATTCTAA-----

barkeriB TTTAAATCAT CGATCATAAT CTAATGATCA ATTCTAA-----

barkeriC CTGTGGATCT CTAGTCTCTC-----

mazeiA TTTGGATCTC TTGTCTCT-----

mazeiB TCCATAT-----

mazeiC TCCATAT-----

acetivoransB TTTGGATCTC CTGTCTCT-----

acetivoransA TTTGGATCTC CTGTCTCT-----

acetivoransC TTTGGATCTC CTGTCTCT-----

301

thermoauto_A -----TTTTTAATG GATTTTCCTT TGGTGCACCC GC-----CCAT TCAGGT-----

thermoauto_B -----TTTTTAATG GATTTTCCTT TGGTGCACCC GC-----CCAT TCAGGT-----

thermophila_A -----TTTTTAATG GATTTTCCTT TGGTGCACCC GT-----

thermophila_B -----TTTTTAATG GATTTTCCTT TGGTGCACCC GT-----

LCMS --GCCTTTGC AAGGCAGAGG CCATGGGTTT GAGTCCCAGC AAGTCCACTT AC---AT- TTTTAAATGC ACCGAGTAA TAATTT-----

stadtmanae_A TGGCACTAAC TAACTAGAG- ---TAGATT AGAAAAAAT AAGTCCATAC AA---TTT- GTATTGATTT CTAATATPAT TAATTTATTC AATTAGTTTG

stadtmanae_B TGGCACTAAC TAACTAGAG- ---TAGATTA GAAAAAAT AAGTCCATAC AA---TTT- TATTGATTTT TGAGTATTAT TAATTTATTC AATTAGTTTG

stadtmanae_C TGGCACTAAC TAACTAGAG- ---TAGATTA GAAAAAAT AAGTCCATAC AA---TTT- TATTGATTTT TGAGTATTAT TAATTTATTC AATTAGTTTG

stadtmanae_D TGGCACTAAC TAACTAGAG- ---TAGATT AGAAAAAAT AAGTCCATAC AATTTTTTTT TATTGATTTT TGAGTATTAT TAATTTATTC AATTAGTTTG

burtonii_A -----TAAGATT CACAATCATC AAGTGCACCC AG-----CAAG TAATGT-----

burtonii_B -----TAAGATT CACAATCATC AAGTGCACCC AG-----CAAG TAATGT-----

burtonii_C -----TAAGATT CACAATCATC AAGTGCACCC AG-----CAAG TAATGT-----

barkeriA -----CTCATC AAATGCACCC GG-----AAAG TAAAT-----

barkeriB -----CTCATC AAATGCACCC GG-----AAAG TAAAT-----

barkeriC -----TATTT TTATGCACCC GG-----AAAG TAAAT-----

mazeiA -----CTCTTT TTGTGCACCC GG-----AAAG TAAAT-----

mazeiB -----TAATCTC ACAAAATCATC AAGTGCACCC GG-----AAAG TAAAT-----

mazeiC -----TAATCTC ACAAAATCATC AAGTGCACCC GG-----AAAG TAAAT-----

acetivoransB -----TCTTT TTGTGCACCC GG-----AAAG TAGTT-----

acetivoransA -----CTATTAATTT TATAATCATC AAGTGCACCC GG-----AAAG TAGTT-----

acetivoransC -----CTATTAATTT TATAATCATC AAGTGCACCC GG-----AAAG TAGTT-----

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thermoauto_A -----TGT GGG----- ---ATGTGGT GGTGAAGTTG GAATGATATG G-----
thermoauto_B -----TGT GGG----- ---ATGTGGT GGTGAAGTTG GAATGATATG G-----
thermophila_A -----GGAGAAC TGCTCCACAG GGAAGGGCTG AT-----
thermophila_B -----GGAGAAC TGCTCCACAG GGAAGGGCTG AT-----
LCMS -----TATTT GGG----- ---AAGGAT A-----GATTTG TTAAATACC CACCCGGTAT TTATGAG--- ---AAATA TTTTCTCTTA
stadtmanae_A TTAGATGTTT GTG-TAATAT ATAATAGAAT AGTAATATTT CTATGGTTTA CTTAAATAAT ATAATAGTAT TAATACGTTT TAATAAGATT TGCTTATATA
stadtmanae_B TTAGATGTTT GTGTAATAAT ATAATAGAAT AGTAATATTT CTATGGTTTA CTTAAATAAT ATAATAGTAT TAATACGTTT TAATAAGATT TGCTTATATA
stadtmanae_C TTAGATGTTT GTGTAATAAT ATAATAGAAT AGTAATATTT --ATGGTTTA CTTAAATAAT ATAATAGTAT TAATACGTTT TAATAAGATT TGCTTATATA
stadtmanae_D TTAGATGTTT GTGTAATAAT ATAATAGAAT AGTAATATTT --ATGGTTTA CTTAAATAAT ATAATAGTAT TAATACGTTT TAATAAGATT TGCTTATATA
burtonii_A -----TGCTT GGG----- ---AAGGAT G-----GATGTG CCTGA-----
burtonii_B -----TGCTT GGG----- ---AAGGAT G-----GATGTG CCTGA-----
burtonii_C -----TGCTT GGG----- ---AAGGAT G-----GATGTG CCTGA-----
barkeriA -----TTC GGG----- ---GAAGGC GGATTGCCTG CGTTGACACG C-----
barkeriB -----TTC GGG----- ---GAAGGC GGATTGCCTG CGTTGACACG C-----
barkeriC -----TTC GGG----- ---GAAGGC GGATTGCCTG CGTTGACACG C-----
mazeiA -----TTC GGG----- ---GAAGGC GGATTGCCTG CGCGGAACCG C-----
mazeiB -----TTC GGG----- ---GAAGGC GGATTGCCTG CGCGGAACCG C-----
mazeiC -----TTC GGG----- ---GAAGGC GGATTGCCTG CGCGGAACCG C-----
acetivoransB -----TTC GGG----- ---GAAGGC GGATTGCCTG TGCTGAAGCT C-----
acetivoransA -----TTC GGG----- ---GAAGGC GGATTGCCTG TGCTGAAGCT C-----
acetivoransC -----TTC GGG----- ---GAAGGC GGATTGCCTG TGCTGAAGCT C-----

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thermoauto_A -----T GATTTCACG CATAGGAGAA ACC-----C GATTGTAATC --CAAAACTG GCATTAACCTG ACCAGAGAGA
thermoauto_B -----T GATTTCACG CATAGGAGAA ACC-----C GATTGTAATC --CAAAACTG GCATTAACCTG ACCAGAGAGA
thermophila_A -----AAT CAAGATGAGG CCA-----C GTATACGCTT TGCAGACCAG ACGCTCACTG -----A
thermophila_B -----AAT CAAGATGAGG CCA-----C GTATACGCTT TGCAGACCAG ACGCTCACTG -----A
LCMS TGTATTTAGC TTTTTCGCTT TTGCATAAAA CAAAGTGAAA TCG-----T GTATATGAAT GGCATATTAG ACGCTCACTG -----A
stadtmanae_A TATTTTTTGC TTTGTTTGAT TTGTTTTTAA TCTACTACAG ACAATATTAT TTTTGTCACT ATATATAATA GGAAAA--AA AAGAACAACCTG -----TATA
stadtmanae_B TATTTTTTGC TTTGTTTGAT TTGTTTTTAA TCTACTACAG ACAATATTAT TTTTGTCACT ATATATAATA GGAAAA--AA AAGAACAACCTG -----TATA
stadtmanae_C TATTTTTTGC TTTGTTTGAT TTGTTTTTAA TCTACTACAG ACAATATTAT TTTTGTCACT ATATATAATA GGAAAA--AA AAGAACAACCTG -----TATA
stadtmanae_D TATTTTTTGC TTTGTTTGAT TTGTTTTTAA TCTACTACAG ACAATATTAT TTTTGTCACT ATATATAATA GGAAAA--AA AAGAACAACCTA -----TGTA
burtonii_A -----TACC CCATATCAGG TACTATGAGA TCA-----T GTATACATAT TACATATCAG ACGCTCACTG -----G
burtonii_B -----TACC CCATATCAGG TACTATGAGA TCA-----T GTATACATAT TACATATCAG ACGCTCACTG -----G
burtonii_C -----TACC CCATATCAGG TACTATGAGA TCA-----T GTATACATAT TACATATCAG ACGCTCACTG -----G
barkeriA -----AGG TACA-TGAAG TCA-----T GTATAAGTGC TGTACTGCG ACGCTTCTG -----G
barkeriB -----AGG TACA-TGAAG TCA-----T GTATAAGTGC TGTACTGCG ACGCTTCTG -----G
barkeriC -----AGG TACA-TGAAG TCA-----T GTATAAGTGC TGTACTGCG ACGCTTCTG -----G
mazeiA -----AGG CACA-TGAAG TCG-----T GTATAGTGC TGTATATTGA ACGCTAACCTG -----G
mazeiB -----AGG CACA-TGAAG TCG-----T GTATAGTGC TGTATATTGA ACGCTAACCTG -----G
mazeiC -----AGG CACA-TGAAG TCG-----T GTATAGTGC TGTATATTGA ACGCTAACCTG -----G
acetivoransB -----AGG CATA-TGAAG TCG-----T GTATATGTGC TGTACTGCG ACGCTTCTG -----G
acetivoransA -----AGG CATA-TGAAG TCG-----T GTATATGTGC TGTACTGCG ACGCTTCTG -----G
acetivoransC -----AGG CATA-TGAAG TCG-----T GTATATGTGC TGTACTGCG ACGCTTCTG -----A

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thermoauto_A AGGC-AGTTA AACCAAACCC TAGCTTA----
thermoauto_B AGGC-AGTTA AACCAAACCC TAGCTTA----
thermophila_A GATC-AGTGG GACGATTAAG CTGCT----
thermophila_B GATC-AGTGG GACGATTAAG CTGCT----
LCMS ATATAAGTGA GAGTGATTCT GGTAC----
stadtmanae_A AAAT-GGTGA AATTTTGTAT AATAAAAAAT TTTTTTCTT----
stadtmanae_B AAAT-GGTGA AATTTTGTAT AATAAAAAAT TTTTTTCTT----
stadtmanae_C AAAT-GGTGA AATTTTGTAT AATAAAAAAT TTTTTTCTT----
stadtmanae_D AAAT-GGTGA AATTTTGTAT AATAAAAAAT TTTTTTCTT----
burtonii_A ACAA-AGTGA GATGGACTCT GGTA----
burtonii_B ACAA-AGTGA GATGGACTCT GGTA----
burtonii_C ACAA-AGTGA GATGGACTCT GGTA----
barkeriA ACCT-GGTGA GGATACACAG GAA-----
barkeriB ACCT-GGTGA GGATACACAG GAA-----
barkeriC ACCT-GGTGA GGATACACAG GAA-----
mazeiA ACCT-GGTGA GGTATATAGG AAT-----
mazeiB ACCT-GGTGA GGTATATAGG AAT-----
mazeiC ACCT-GGTGA GGTATATAGG AA-----
acetivoransB ACCT-GGTGA GGTAAATAGG AATTATGCTA TCAGGTGGAT GGCTCGGCTC AAGAGCTTA
acetivoransA ACCT-GGTGA GGTAAATAGG AATTATGCTA TCAGGTGGAT GGCTCGGCTC AAGAGCTTA
acetivoransC AAGT-AGTT- -----

```

Figure S1. The LCMS ITS region encodes a tRNA and shows sequence similarity to the ITS regions of several methanogens. The alignment includes sequences from: *Methanosaeta thermophila* (NC_008553), *Methanosarcina barkeri* (NC_007349), *Ms. acetivorans* (NC_003552), *Ms. mazei* (NC_003901), *Methanococcoides burtonii* (NC_007955), *Methanobacterium thermoautotrophicus* (NC_000916), *Methanosphaera stadtmanae* (NC_007681), and Lost City Methanosarcinales (GQ273207).

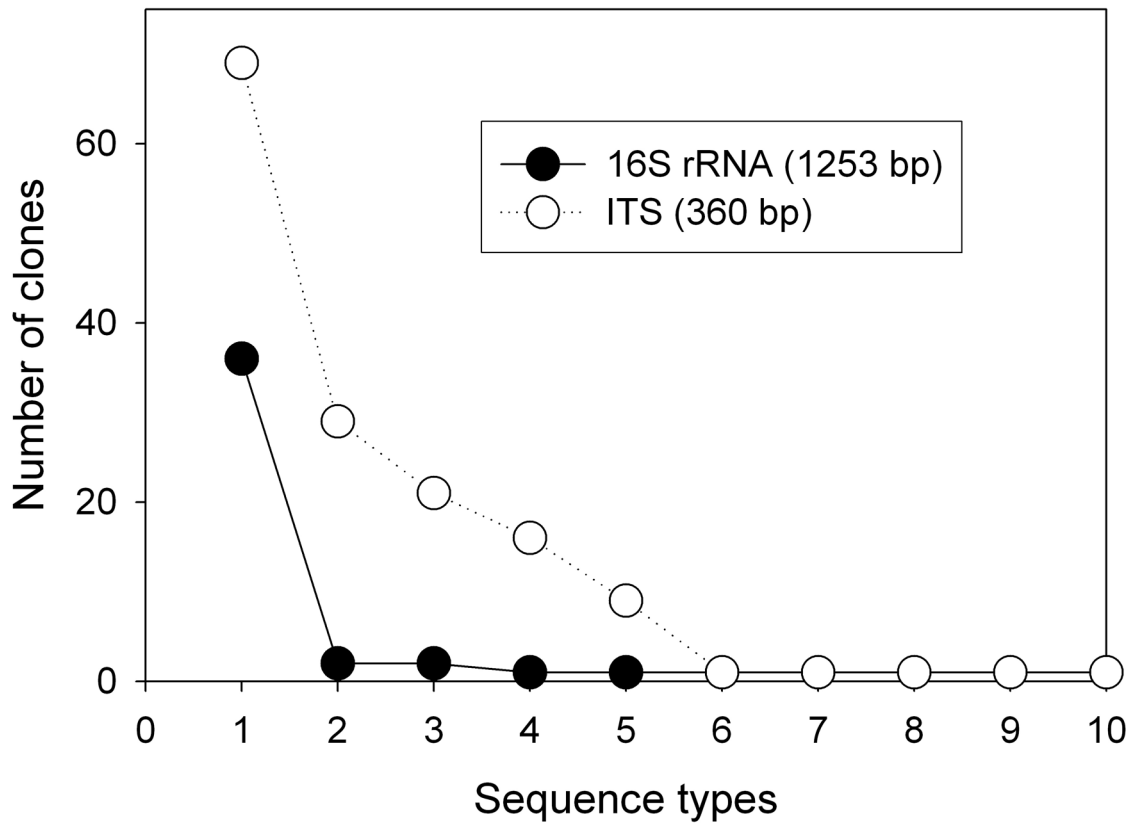


Figure S2. Rank-abundance plot showing the number of clones sharing the 10 most frequently occurring 16S rRNA and ITS sequences in samples LC0424 and LC1408, both of which were collected from the Poseidon chimney (Marker 3). Only one 16S rRNA sequence occurs more than twice, but five ITS sequences occur many times in this sample. As shown in Figure 3b, other samples contain different abundant ITS sequences.